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(FILE 'HOME' ENTERED AT 15:15:54 ON 12 JAN 2005)

FILE 'MEDLINE, AGRICOLA, SCISEARCH, CAPLUS, MEDICONF' ENTERED AT 15:16:09 ON 12 JAN 2005

L1 95924 S OOPLASTOID OR OOCYTES
L2 116795 S OOPLASTOID OR OOCYTE
L3 24464 S ENUCLEAT? OR REMOV?(5W)NUCLE?
L4 3847 S METAPHASE(1W)II
L5 10542 S ZONA PELLUCIDA
L6 1438 S L2(L)L3
L7 27 S L2(L)L3(L)L5
L8 8 S L2(L)L3(L)L5(L)L4
L9 4 DUP REM L8 (4 DUPLICATES REMOVED)
L10 17 DUP REM L7 (10 DUPLICATES REMOVED)
L11 17 SORT L10 PY
E LEVANDUSKI M?/AU
L12 6 S E2
L13 1 S E4
L14 7 S L12 OR L13
L15 4 S L14 AND L2
L16 2 DUP REM L15 (2 DUPLICATES REMOVED)

=> d an ti so au ab pi 116 1-2

L16 ANSWER 1 OF 2 CAPLUS COPYRIGHT 2005 ACS on STN
AN 2003:473152 CAPLUS
DN 139:19332
TI Pluripotent stem cells derived without the use of embryos or fetal tissue
SO U.S. Pat. Appl. Publ., 37 pp.
CODEN: USXXCO
IN Levanduski, Mike
AB This invention provides a method for deriving precursors to pluripotent non-embryonic stem (P-PNES) and pluripotent non-embryonic stem (PNES) cell lines. The present invention involves nuclear transfer of genetic material from a somatic cell into an enucleated, zona pellucida free human ooplastoid having a reduced amount of total cytoplasm. The present invention provides a new source for obtaining human and other animal pluripotent stem cells. The source utilizes as starting materials an oocyte and a somatic cell as the starting materials but does not require the use, creation and/or destruction of embryos or fetal tissue and does not in any way involve creating a cloned being. The oocyte never becomes fertilized and never develops into an embryo. Rather, portions of the oocyte cytoplasm are extracted and combined with the nuclear material of individual mature somatic cells in a manner that precludes embryo formation. Murine, bovine, and human examples of the procedure are demonstrated. Subsequently, the newly constructed P-PNES cells are cultured in vitro and give rise to PNES cells and cell colonies. Methods are described for culturing the P-PNES cells to yield purified PNES cells which have the ability to differentiate into cells derived from mesoderm, endoderm, and ectoderm germ layers. Methods are described for maintaining and proliferating PNES cells in culture in an undifferentiated state. Methods and results are described for anal. and validation of pluripotency of PNES cells including cell morphol., cell surface markers, pluripotent tumor development in SCID mouse, karyotyping, immortality in in vitro culture.

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI US 2003113910	A1	20030619	US 2001-26420	20011219
WO 2003052080	A2	20030626	WO 2002-US40562	20021218
WO 2003052080	A3	20031127		

W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW

RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AM, AZ, BY,
 KG, KZ, MD, RU, TJ, TM, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES,
 FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, SI, SK, TR, BF, BJ,
 CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG
 EP 1465992 A2 20041013 EP 2002-805209 20021218
 R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,
 IE, SI, LT, LV, FI, RO, MK, CY, AL, TR, BG, CZ, EE, SK

L16 ANSWER 2 OF 2 MEDLINE on STN DUPLICATE 1
 AN 92389260 MEDLINE
 TI Viable embryos and normal calves after nuclear transfer into Hoechst
 stained enucleated demi-oocytes of cows.
 SO Journal of reproduction and fertility, (1992 Jul) 95 (2) 475-80.
 Journal code: 0376367. ISSN: 0022-4251.
 AU Westhusin M E; Levanduski M J; Scarborough R; Looney C R;
 Bondioli K R
 AB Bovine oocytes were bisected, stained with Hoechst 33342 and
 observed under a fluorescent microscope to identify nucleated and
 enucleated demi-oocytes. Other oocytes were bisected
 but not stained, or bisected and only half of each oocyte
 stained, and viewed under a fluorescent microscope. The oocytes
 were then used for nuclear transfer by fusing them with embryonic
 blastomeres from a 5-6 day bovine embryo. The fusion rate and proportion
 developing into compact morulae or blastocysts was compared among
 different types of demi-oocytes. Expt 1 examined the effect of
 staining and indicated no effect on either fusion rate or embryonic
 development whether or not the oocytes were stained. In Expt 2,
 stained and unstained nucleated and enucleated oocytes were
 compared. As in the first experiment, there were no differences between
 stained and unstained demi-oocytes. There was no difference
 between fusion rates of nucleated and enucleated oocytes.
 However, there was a significant difference in embryonic development
 between nucleated (10.4%) and enucleated (22.6%) demi-oocytes (P
 less than 0.05). In a final experiment, stained and unstained enucleated
 oocytes were used for nuclear transfer and the resulting embryos
 transferred into recipient cows. There was no difference in pregnancy
 rates or in the number of normal calves born whether stained or unstained
 recipient oocytes were used. Results from these experiments
 indicate that Hoechst staining and fluorescent microscopy can be used to
 identify enucleated demi-oocytes, and that these can be used for
 nuclear transfer, and result in viable embryos and normal calves. (ABSTRACT
 TRUNCATED AT 250 WORDS)